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# Mating system and *avpr1a* promoter variation in primates

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It has been suggested that primate mating and social behaviours may be influenced by variation in promoter region repetitive DNA of the vasopressin receptor 1a gene (*avpr1a*). We show that male mating behaviour does not covary in a simple way with promoter repetitive DNA in 12 Old World primates. We found that one microsatellite (–553 bp upstream) was present in all species, irrespective of their behaviour. By contrast, two microsatellites (–3956 and –3625 bp upstream) were present only in some species, yet this variation did not correlate with behaviour. These findings agree with a recent comparative analysis of voles and show that the variation in repetitive DNA in the *avpr1a* promoter region does not generally explain variation in male mating behaviour. Phylogenetic analysis revealed a GAGTA motif that has been independently deleted three times and involved in another larger deletion. Importantly, the presence/absence of this GAGTA motif leads to changes in predicted transcription factor-binding sites. Given the repeated loss of this motif, we speculate that it might be of functional relevance. We suggest that such non-repetitive variation, either in indels or in sequence variation, are likely to be important in explaining interspecific variation in *avpr1a* expression.

**Keywords:** primates; mating behaviour; microsatellites; vasopressin; *avpr1a*; voles

## 1. INTRODUCTION

There is great interest in how much variation in social and mating behaviours is caused by genetic variation (Robinson *et al.* 2005), particularly by genes of large effect. Possibly, the best example is the vasopressin receptor 1a gene (*avpr1a*), which codes for the receptor (AVPR1A) of the nonapeptide arginine vasopressin. This gene has been shown in a series of elegant studies to have a major role explaining both inter- and intraspecific differences in the social and mating behaviours of voles (Young *et al.* 1997, 1999; Lim *et al.* 2004; Hammock & Young 2005). Data suggest that differences in receptor distribution and behaviour are caused by variation in repetitive DNA approximately 500 bp upstream of the *avpr1a* transcription start site

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(Hammock & Young 2005). In monogamous ‘social’ vole species (prairie and pine voles), where males pair bond with their mating partners, there are larger blocks of repetitive DNA upstream of the transcription start site compared with polygamous ‘asocial’ vole species (e.g. montane and meadow voles), where males do not pair bond.

Hammock & Young (2005) recently suggested that, as in voles, differences in repetitive non-coding DNA may explain behavioural differences among primate species. This suggestion was motivated by their discovery that humans and bonobos (*Pan paniscus*) differ from chimpanzees (*Pan troglodytes*) in the number of microsatellites in a non-coding region upstream of *avpr1a*. Humans and bonobos have two microsatellites (–3956 and –3625 bp upstream of the start site; Thibonnier *et al.* 2000), whereas a 360 bp region, including the –3625 bp microsatellite, is absent in the chimpanzee (*P. troglodytes*). However, testing this idea needs data from more species with well-defined differences in social and mating behaviours and known phylogenetic relationships. Furthermore, the –3956 and –3625 bp microsatellites are both dinucleotide repeats and are much further upstream from the transcription start site than the tetranucleotide repeat microsatellite that influences mating and social behaviours in voles (Fink *et al.* 2006). There is a third microsatellite, not considered by Hammock and Young, 553 bp upstream of *avpr1a*, which has the same repeat type (tetranucleotide) and is in a similar position to the vole microsatellite (Thibonnier *et al.* 2000). *A priori*, the –553 bp microsatellite is perhaps a better candidate than either the –3956 or the –3625 bp microsatellite for influencing male behaviour.

We have expanded upon Hammock & Young’s speculations by comparing the structure of all three microsatellites (–553, –3625 and –3956 bp) in a sample of 12 primates, comprising humans, five great apes (bonobo, two subspecies of chimpanzee, gorilla and orang-utan), three species of gibbon (lar, siamang and crested) and three Old World monkeys (green monkey, hamadryas baboon and rhesus macaque). Importantly, these species show great variation in the mating behaviour of males, particularly the duration and exclusivity of male–female post-mating affiliations, and the main topology of the phylogeny is known without ambiguity (except gibbon relationships). Male humans and gibbons are capable of forming long-term pair bonds with female mates, whereas male chimpanzees, bonobos, macaques and green monkeys show only weak non-exclusive associations with individual female mates. Male hamadryas baboons and gorillas have strong, but non-exclusive, associations with female mates while orang-utans form brief consortships (see electronic supplementary material for support for these classifications). Given this variation in social behaviour, we asked the following questions:

- (i) does variation in any of the microsatellites upstream of *avpr1a* covary with social behaviour in a predictable way? For example, do pair-bonding gibbons and humans have larger regions of repetitive DNA than species that do not pair bond? and
- (ii) can we identify, using a phylogenetic approach, regions other than the microsatellites that may be functionally important?

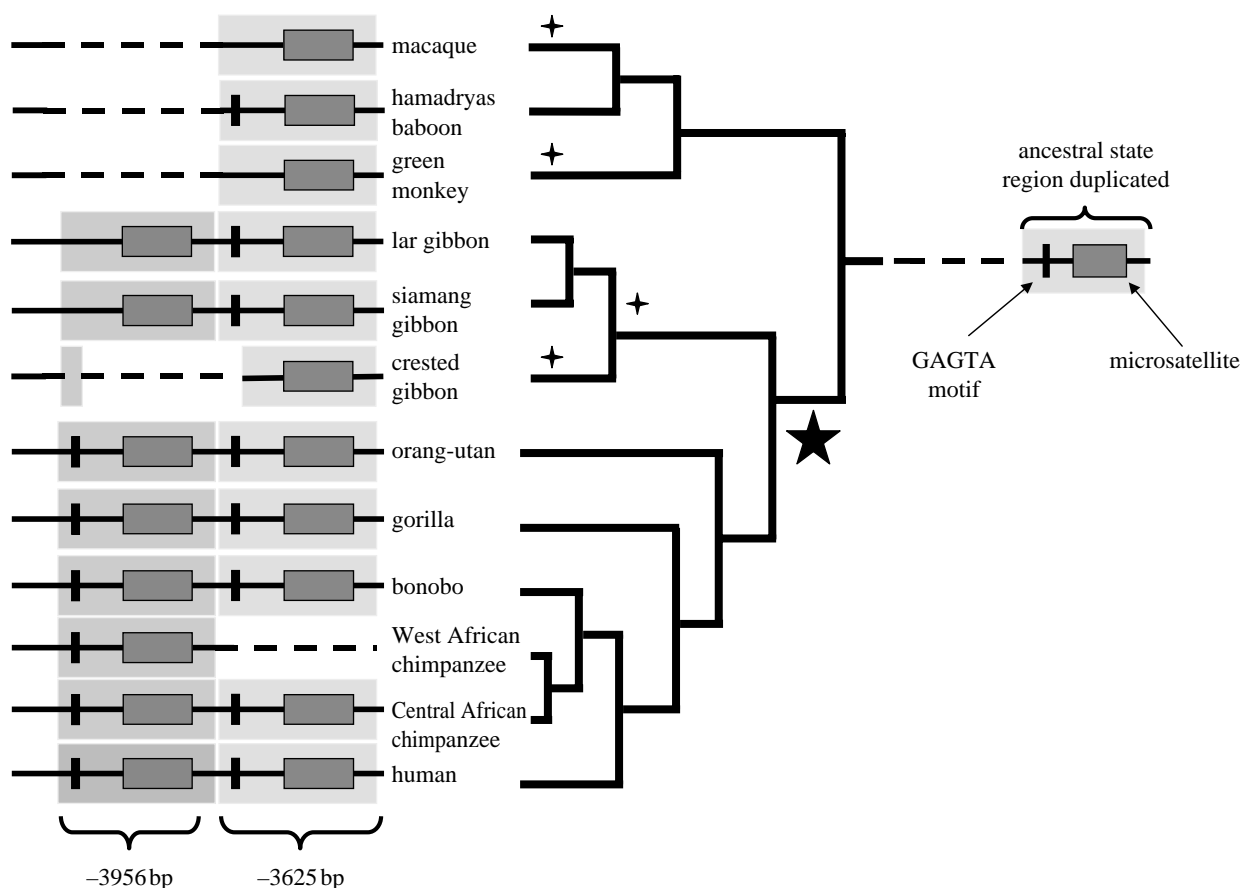


Figure 1. *avpr1a* dinucleotide microsatellite structure mapped onto the phylogeny for 12 Old World primates. Shaded rectangles labelled  $-3956$  and  $-3625$  bp show duplicated regions containing microsatellites (dark-shaded rectangles) and the GAGTA motif (black vertical bars). Broken lines indicate sequence absences either owing to deletions or because they are basal to the  $-3625$  bp duplication event. Five pointed star shows position of the  $-3625$  bp duplication that gave rise to the  $-3956$  bp region, four pointed stars show losses of GAGTA motif.

## 2. MATERIAL AND METHODS

### (a) Sequences

From a sample of 12 Old World primate taxa (see electronic supplementary material), we amplified and sequenced two regions upstream of the *avpr1a* transcription start site that contained up to three repetitive DNA elements. We amplified and sequenced the 'dinucleotide region' (including  $-3956$  and  $-3625$  bp dinucleotide microsatellites, Thibonnier *et al.* 2000) using primers (5' *Cla*I site removed) described in Hammock & Young (2005), and the 'tetranucleotide region' ( $-553$  bp tetranucleotide microsatellite) using primers from Kim *et al.* (2002). PCR products were cloned into pGEM Easy T (Promega) plasmids. We sequenced three to six clones per individual in both directions.

### (b) Alignments

We arbitrarily chose the allele with the longest repeat at the  $-3625$  bp microsatellite (except in the West African chimpanzee where we chose the longest  $-3956$  bp microsatellite allele) and we aligned all sequences by eye.

### (c) Identification of transcription factor-binding sites

Mapping aligned sequences on to the phylogeny identified a number of insertion/deletion events that may have caused changes in transcription factor-binding sites. We identified potential transcription factor-binding sites using the program ALIBABA v. 2.1 (Grabe 2000, <http://www.gene-regulation.com/pub/programs/alibaba2/index.htm>) to search the Transfac v. 4.0 database (Wingender *et al.* 2000).

## 3. RESULTS

We found considerable variation among species in the  $-3956$  and  $-3625$  bp repeat regions (figure S1 in the electronic supplementary material). The three most basal species in the phylogeny (hamadryas baboon,

macaque and green monkey) had the  $-3625$  bp repeat region, but lacked the  $-3956$  bp region (figure 1). In seven out of nine ape taxa, both the  $-3956$  and  $-3625$  bp microsatellites were present, but the  $-3956$  bp repeat region was missing in the black-crested gibbon and the  $-3625$  bp repeat region was missing in the West African chimpanzee (Hammock & Young 2005; figure 1). Interestingly, both the  $-3956$  and  $-3625$  bp microsatellites were present in chimpanzees from Central Africa, showing that chimpanzees are polymorphic for the deletion of the  $-3625$  bp microsatellite. This pattern of change, mapped onto the phylogeny, suggests that the  $-3956$  bp microsatellite arose by tandem duplication of the  $-3625$  bp region in the ancestor to all apes (figure 1). Subsequently, there have been two independent losses of a repeat region in the black-crested gibbon and in West African chimpanzees. In contrast to the evolutionary lability of the  $-3956$  and  $-3625$  bp repeat regions, the  $-553$  bp tetranucleotide microsatellite was present in all 12 taxa (figure S2 in the electronic supplementary material).

Mapping indels onto the phylogeny suggested that non-repetitive regions of the *avpr1a* promoter may be functionally important. For example, before the  $-3956$  and  $-3625$  bp microsatellites, there is a GAGTA motif that is either present or absent (figure S1 in the electronic supplementary material: positions 94–98 and 445–449). All great apes, except the western

chimpanzee, have two GAGTA motifs, a pattern that suggests that the ancestral state was hamadryas-like. Given this, there have been at least three independent losses of the GAGTA motif: in macaques, in green monkeys, and in the ancestor to gibbons. In contrast to these repeated losses, the GAGTA motif is embedded in a 35–40 bp block of sequence (figure S1 in the electronic supplementary material: positions 76–110 and 427–466) that is conserved, with only single substitutions separating all gibbons (figure S1 in the electronic supplementary material: –3956 bp region: position 79), or the siamang (figure S1 in the electronic supplementary material: –3625 bp region: position 454), from all other taxa.

#### 4. DISCUSSION

The variation in the presence/absence of dinucleotide repeat regions (–3956 and –3625 bp) does not covary in a simple way with male behaviour (figure 1). For example, differences in repeat structure between chimpanzee subspecies and among gibbons do not correspond to large differences in male behaviour. Likewise, species with dissimilar male behaviours (e.g. orang-utans, bonobos and lar/symphalangus gibbons) have the same repeat region structure. We also found no evidence that variation in repetitive DNA approximately 500 bp upstream of the transcription start site explained variation in male mating behaviour, as the –553 bp tetranucleotide repeat region was present in all taxa irrespective of their behaviour (figure S2 in the electronic supplementary material). Furthermore, our prediction that monogamous gibbons would have more repetitive DNA than non-monogamous taxa was not supported by our data. It might be argued that all primates studied here are capable of forming strong social bonds in contrast to voles where males are either able (e.g. prairie and pine voles) or unable (e.g. montane and meadow voles) to do so. This re-categorization of the behaviour of the primates still leads to the conclusion that variation in repetitive DNA in the –3956 and –3625 bp microsatellite regions does not covary simply with behaviour, as the lack of variation in behaviour (all primates can form social bonds) would be at odds with the high variation in promoter repetitive DNA we found. We suggest, therefore, that our findings agree with a recent study that found no general relationship between *avpr1a* repeat structure and male mating behaviour across a large number of *Microtus* voles (Fink *et al.* 2006).

Although our results, and those of Fink *et al.* (2006), clearly show no association of repeat structure and behaviour, it is important to stress that neither our study, nor that of Fink *et al.* (2006), tested whether AVPR1A receptor distribution in the brains of rodents and primates covaries with behaviour. This is still a viable hypothesis that needs to be tested in both voles and primates (Young & Hammock 2007). It is, therefore, premature to conclude that there is no general relationship between the gene *avpr1a* and male behaviour.

One possibility is that non-repetitive, as well as repetitive, regions in the promoter influence the expression of *avpr1a* (Hammock & Young 2002). For example, our phylogenetic analysis showed the repeated

loss of the same GAGTA motif and the possible role of the GAGTA repeat in the large deletion in the black-crested gibbon (the –3956 bp repeat region deletion spans the two GAGTA motifs). Although repeated losses might be interpreted as a lack of conservation, the repeated losses of the GAGTA motif contrasts with the conservation of the surrounding 40 bp, suggesting that this region might have some functional role in the expression of *avpr1a*. In support of this notion, searches of the Transfac database (Wingender *et al.* 2000) with the program ALIBABA v. 2.1 (Grabe 2000) show that the loss of the GAGTA motif alters predicted transcription factor-binding sites, for example, changing *HNF-3b* to *MEF2* (figure S3 in the electronic supplementary material), with the latter being important in neuronal development and survival (Shalizi & Bonni 2005). Additionally, there are other indels and single nucleotide variation (see electronic supplementary material) that may be important in regulating cell-type-dependent expression of *avpr1a*.

In conclusion, it appears that there is no simple relationship between *avpr1a* promoter region repetitive DNA and male mating behaviour. This does not mean, however, that variation in expression of *avpr1a* is necessarily unimportant in explaining interspecific differences in mammalian mating/social behaviours, but that the regulation of expression is more complex than previously thought (Young & Hammock 2007).

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